

Effect of supplementation with methyl-donor nutrients on neurodevelopment and cognition: considerations for future research

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Pregnancy represents a critical period in fetal development, such that the prenatal environment can, in part, establish a lifelong trajectory of health or disease for the offspring. Poor nutrition (macro- or micronutrient deficiencies) can adversely affect brain development and significantly increase offspring risk for metabolic and neurological disease development. The concentration of dietary methyl-donor nutrients is known to alter DNA methylation in the brain, and alterations in DNA methylation can have long-lasting effects on gene expression and neuronal function. The decreased availability of methyl-donor nutrients to the developing fetus in models of poor maternal nutrition is one mechanism hypothesized to link maternal malnutrition and disease risk in offspring. Animal studies indicate that supplementation of both maternal and postnatal (early- and later-life) diets with methyl-donor nutrients can attenuate disease risk in offspring; however, clinical research is more equivocal. The objective of this review is to summarize how specific methyl-donor nutrient deficiencies and excesses during pre- and postnatal life alter neurodevelopment and cognition. Emphasis is placed on reviewing the current literature, highlighting challenges within nutrient supplementation research, and considering potential strategies to ensure robust findings in future studies.

INTRODUCTION

Pregnancy and early life are critical periods of development, particularly with regard to neurocognitive endpoints, because significant brain maturation continues throughout the postnatal period. Beginning in pregnancy but continuing well into adolescence, neural systems, particularly those in the prefrontal cortex that are essential for cognition and executive function processes such as attention and cognitive flexibility, are established and refined. This significant postnatal development means that the neural systems underlying cognition remain vulnerable to adverse environmental influences, such as poor diet, alcohol or drugs of abuse, stressors, or inflammation, for a prolonged period of

time. Early exposure to these environmental challenges can therefore increase the risk for a wide range of adverse neurodevelopmental outcomes, such as cognitive deficits, executive function deficits, and other social and emotional behavioral problems.^{1,2}

Epigenetic modifications and their resultant changes in gene expression have been proposed as one mechanism linking poor early-life environment with long-term changes in physiology and increased disease risk. Epigenetics, meaning “on” or “above” the genome, is the study of modifications to DNA or histones that can regulate gene expression, some of which can be heritable and/or reversible.³ In the context of poor early-life nutrition, the decreased availability of methyl-donor nutrients to the developing offspring is one mechanism

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hypothesized to link maternal malnutrition and disease risk in offspring. Animal studies indicate that supplementation of both maternal and postnatal (early- and later-life) diets with methyl-donor nutrients can attenuate disease risk in offspring. This review will first examine 1-carbon metabolism and methyl-donor nutrients and will then summarize the literature pertaining to effects of methyl-donor supplementation on cognitive endpoints in both animals and humans. It will conclude with a discussion of some of the caveats and limitations within the field and will consider opportunities for future studies to fill gaps in the current knowledge.

ONE-CARBON METABOLISM AND METHYL-DONOR NUTRIENTS

Methyltransferases receive methyl groups to perform methylation reactions through the conversion of S-adenosylmethionine to S-adenosylhomocysteine. This reaction is one in a cyclical series of biochemical reactions called 1-carbon metabolism (Figure 1). One-carbon metabolism refers to the intersection of folate and methionine metabolism cycles that donate and regenerate 1-carbon units for various reactions in the cell.^{4–6} Through these reactions, cells synthesize purines, thymidylate, creatine, phosphatidylcholine, and multiple hormones. One-carbon metabolism also provides methyl groups for at least 50 different methylation reactions that occur within the cell, including methylation of RNA, DNA, histones, neurotransmitters, and other proteins. It is also involved in the catabolism of choline and histidine, the interconversion of serine and glycine, and in almost every cellular process; thus, it is important that these reactions occur efficiently.

One-carbon metabolism is highly dependent on nutritional status. In Figure 1, highlighted in gray are nutrients essential for 1-carbon metabolism, known as methyl donors. Vitamin B₁₂, vitamin B₆, vitamin B₉ (folate), methionine, betaine, choline, and zinc are all necessary for 1-carbon metabolism. These methyl-donor nutrients are critical intermediates or cofactors for enzymes involved in 1-carbon metabolism. Specifically within the brain, methyl-donor nutrients are known to be involved in multiple neurotransmitter systems, epigenetic modifications, and cellular membrane structures. For example, choline is a precursor for acetylcholine synthesis, a major neurotransmitter system involved in attention, memory, and motivation.^{7–9} It is also a precursor for phosphatidylcholine, a class of phospholipids integral to the formation of biological membranes.¹⁰ Folate is integral to the donation of methyl groups that are utilized by multiple methyltransferases.¹¹ DNA methyltransferases mediate the addition of methyl groups to DNA, a well-studied epigenetic

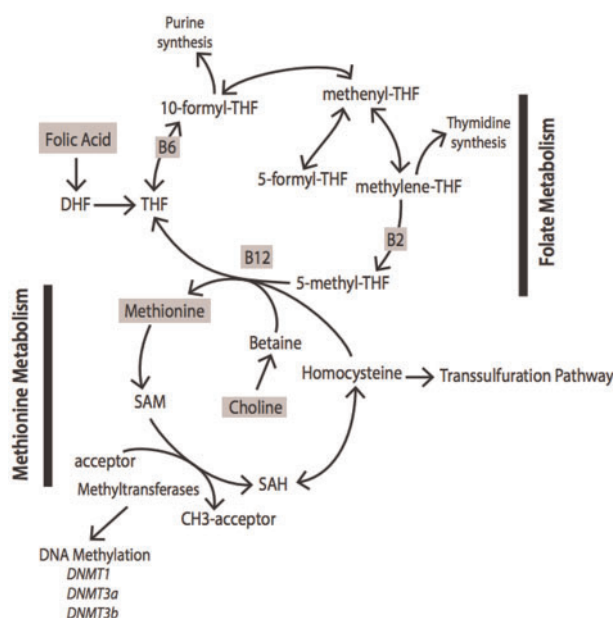


Figure 1 Simplified schematic diagram of the intermediates, enzymes, and nutrients involved in 1-carbon metabolism. Abbreviations: DHF, dihydrofolate; SAH, S-adenosylhomocysteine; SAM, S-adenosylmethionine; THF, tetrahydrofolate.

modification known to mediate gene expression changes and neuronal function. Catechol-O-methyltransferase, another methyltransferase, utilizes methyl groups to aid in the degradation of catecholamines (dopamine, epinephrine, norepinephrine). Imbalances in methyl donors can also negatively affect methyltransferase function, S-adenosylmethionine regeneration, and DNA methylation.^{12–17} Additionally, deficiencies and excesses of these nutrients also can alter cognition, behavior, and risk for disease.^{18–24}

Folate metabolism serves to create and transfer 1-carbon units for the biosynthetic processes listed above. Natural folates in the body and diet are typically found in reduced form, mainly 5-methyltetrahydrofolate in humans.²⁵ The tetrahydrofolate backbone, which is the active form of folate, is used to carry and transfer 1-carbon units. One-carbon units, also known as methyl groups, are covalently bound to the 5- and 10-positions on the pteridine ring of tetrahydrofolate. Tetrahydrofolate can be maintained in 3 oxidative states, each of which plays a specific role in biosynthesis (recently reviewed by Ducker and Rabinowitz⁵). Briefly, 5,10-methylenetetrahydrofolate produces thymidine and serine, 5-methyltetrahydrofolate produces methionine, and 10-formyltetrahydrofolate produce purines, formate, and carbon dioxide.

The most abundant form of folate, 5-methyltetrahydrofolate, can be converted to methionine via a vitamin B₁₂-dependent methyltransferase in which the methyl group from 5-methyltetrahydrofolate is transferred

to homocysteine, resulting in the formation of methionine and unsubstituted tetrahydrofolate. Tetrahydrofolate can then begin the folate cycle again, and methionine can continue through its unique metabolism. Methionine is the precursor of S-adenosylmethionine, the universal methyl donor, as it is the final carrier of methyl groups before they are donated to the methyltransferases. Donation of a methyl group from S-adenosylmethionine creates S-adenosylhomocysteine, which is reversibly metabolized into homocysteine. Homocysteine can be remethylated to reform methionine via betaine-homocysteine methyltransferase (expressed in liver and kidney, but not found in the brain or other tissues) or methionine synthase, a vitamin B₁₂-dependent methyltransferase, to recycle through methionine metabolism. Alternatively, homocysteine can enter the transsulfuration pathway, which is required to support glutathione synthesis and remove reactive oxygen species from the brain.²⁶

Methyl-donor nutrients are necessary for normal growth and development of the central nervous system (CNS). Deficiencies in methyl donors and imbalanced concentrations of methyl donors have both been linked to abnormal CNS development as well as to neurological diseases. For example, choline deficiency during gestation alters global and gene-specific DNA methylation in the developing mouse hippocampus, a region important for learning and memory.¹⁷ Increased homocysteine is a potential risk factor for cognitive impairment and other neurological diseases.^{27,28} Additionally, low vitamin B₁₂ status has also been associated with poorer memory performance and reduced microstructural integrity of the hippocampus.²⁹ Conversely, higher folate intake has been associated with lower risk for dementia,³⁰ and it has been suggested that using B vitamins to help remethylate homocysteine to methionine could be one mechanism to improve cognition or to slow cognitive decline.³¹

The efficiency and reactions of 1-carbon metabolism have been shown to affect a number of CNS outcomes, including neural tube closure,^{32,33} neurological diseases,^{34,35} and cognition.³⁶ The efficiency of this cycle is highly dependent on nutritional status, and studies in both animals and humans suggest that nutritional interventions can be used for therapeutic benefit. The subsequent sections will summarize animal studies (Table 1)^{37–50} and human clinical trials (Table 2)^{23,30,31,51–72} linking methyl-donor supplementation with neurodevelopment and cognition.

METHYL DONORS, NEURODEVELOPMENT, AND COGNITION

Beginning at conception, methyl-donor nutrients are important for the development of the CNS, specifically for closure of the neural tube. Beyond this, emerging

evidence suggests that maternal folate status throughout pregnancy affects offspring neurodevelopment, postnatal behavior, and cognition.³⁴ Folate concentrations may decline during pregnancy because of increased demand, larger blood volume, decreased intestinal absorption, or inadequate intake.⁷³ The demand for energy and nutrients required for continuous fetal development also increases during pregnancy. Fortification of grain products with folic acid, a stable oxidized form of folate, was introduced in the United States in the 1990s to help increase the amount of folate consumed by women of childbearing age to prevent the risk of neural tube and other midline defects. Since the introduction of folic acid fortification, the incidence of neural tube defects has decreased by about 25%.³² The use of supplementation with folic acid and other methyl-donor nutrients (mainly vitamin B₁₂, choline, and methionine) as therapeutic interventions to enhance CNS development, prevent neurological disease, and slow cognitive decline has burgeoned in the last 20 years.

Maternal supplementation during pregnancy or lactation

Maternal choline availability influences offspring brain development and function (reviewed by Zeisel and Niculescu¹⁰). In animal models, the positive relationship between maternal choline supplementation and neurodevelopment and cognition is well established. Maternal choline supplementation enhances offspring cognition (fewer working memory errors, fewer responses to reach criterion, fewer reference memory errors, faster behavior acquisition, and shorter escape latency in the Morris water maze), prevents age-related memory decline, enhances hippocampal function,^{40,74} reduces the effects of fetal alcohol exposure in the offspring,^{37,38} increases levels of nerve growth factor in the hippocampus and frontal cortex,⁷⁵ and normalizes choline deficiency-induced DNA hypermethylation.⁷⁶ Choline supplementation can also reverse both the effect of folate deficiency on neurogenesis and apoptosis at E17 in the fetal forebrain⁷⁷ and iron deficiency-induced reprogramming within the hippocampus.⁷⁸ In a mouse model of Down syndrome, maternal supplementation with choline attenuated degeneration of the basal forebrain cholinergic neuron system,⁷⁹ while in a mouse model of autism, high maternal choline consumption improved anxiety-like behaviors and increased social interaction⁴¹ (for review, see Jiang et al⁸⁰). Because of the beneficial findings in rodent models, effects in humans are being investigated. The current results are mixed. Choline levels in early second-trimester maternal plasma are positively correlated with early cognitive development⁵¹ as well as with cognitive

Table 1 Animal studies linking methyl-donor supplementation with neurodevelopment and cognition

Reference	Behavior	Supplementation period	Nutrient(s) supplemented	Outcomes
Thomas et al. (2009) ³⁷	Motor coordination and balance, open field test, spatial learning, working memory	Pregnancy	Choline	Choline supplementation reduced effects of pre-natal ethanol exposure; increased brain weight; and normalized righting reflex, motor coordination, and negative geotaxis
Thomas et al. (2010) ³⁸	Spontaneous alternation, parallel bar motor coordination, Morris water maze, spatial working memory	Pregnancy	Choline	Choline supplementation may reduce severity of fetal alcohol effects. It increased rates of spontaneous alternation; increased memory retention
Naninck et al. (2016) ³⁹	Elevated plus maze, object recognition task, object location task, Morris water maze, and T maze	Pregnancy	Folic acid, vitamin B ₁₂ , vitamin B ₆ , choline, betaine, zinc, and methionine	Supplementation normalized object recognition but did not change object location or performance on Morris water maze
Zhu et al. (2016) ⁴⁰	Barnes maze test	Pregnancy	Choline	Choline supplementation decreased spatial learning deficits
Langley et al. (2015) ⁴¹	Open field test, elevated plus maze, marble burying test	Pregnancy + lactation	Choline	Choline supplementation lowered anxiety levels
Carlin et al. (2013) ⁴²	Fat and sucrose preference	Pregnancy + lactation	Folic acid, vitamin B ₁₂ , choline, betaine, zinc, and methionine	Supplementation decreased palatable food preference
Bahous et al. (2017) ⁴³	Open field test, ladder beam test, novel object recognition test, Y-maze	Pregnancy + lactation	Folic acid	Excessive folic acid supplementation impaired short-term memory; no difference observed in movement score, anxiety, or spatial memory
McKee et al. (2017) ⁴⁴	5-choice serial reaction time task	Early life (3–6 wk)	Folic acid, vitamin B ₁₂ , choline, betaine, zinc, and methionine	Methyl-donor supplementation increased motivation, enhanced operant learning, and shortened reaction time needed to perform correct responses
Carletti et al. (2012) ⁴⁵	Open field test and inhibitory avoidance task	Early life (PD7–30)	Folic acid	Folic acid supplementation reversed neonatal hypoxia-ischemia-induced anxiogenic effects and impaired aversive memory
Sittig et al. (2012) ⁴⁶	Morris water maze	Early life (PD30–60)	Folic acid	Excessive folic acid led to deficits in motivation and spatial memory
Paternain et al. (2016) ⁴⁷	Open field test, forced swim test, novel object recognition task	Adult (8–26 wk)	Folic acid, vitamin B ₁₂ , betaine, and choline	Methyl-donor supplementation reversed early-life-induced depression in rats
Singh et al. (2011) ⁴⁸	Active avoidance, passive avoidance, and elevated plus maze	Adult (6, 11, 16 mo)	Folic acid	Folic acid supplementation for 8 wk improved memory in behavioral tasks

(continued)

Table 1 Continued

Reference	Behavior	Supplementation period	Nutrient(s) supplemented	Outcomes
Brocardo et al. (2010) ⁴⁹	Open field test	Adult (3–4 mo)	Folic acid	Folic acid supplementation decreased hyperlocomotion in a rodent model of bipolar disorder
McKee et al. (2017) ⁴⁴	Progressive ratio task	Adult (32 wk)	Folic acid, vitamin B ₁₂ , choline, betaine, zinc, and methionine	Methyl-donor supplementation increased motivation
Barua et al. (2014) ⁵⁰	Ultrasonic vocalization (P2, P4, P6), social approach task, trace fear conditioning, open field test (2–8 mo)	Pregnancy, lactation, and postweaning	Folic acid	Folic acid supplementation during both gestational development and postweaning period increased anxiety-like behavior, ultrasonic vocalization, and hyperactivity

Abbreviation: PD, postnatal day.

development at 7 years.²³ However, neither supplementing with phosphatidylcholine⁵⁹ nor maternal cord blood choline concentrations⁵³ correlated with enhanced infant cognition (for review, see McCann and Ames⁷⁴ and Jiang et al¹⁸⁰).

In contrast to the link between maternal choline supplementation and neurodevelopment, the positive link between maternal folic acid supplementation and neurodevelopment and cognition is stronger in humans than it is in animal models. In humans, maternal folic acid supplementation is associated with improved verbal, motor, and verbal-executive scores in 4-year-olds.⁵⁴ Maternal use of folic acid supplements is associated with reduced risk of severe language delay in children at 3 years of age.⁵⁵ Higher plasma folate during week 30 of pregnancy is associated with increases in learning, long-term storage, and memory retrieval and a decrease in inattention at 10 years of age.⁵⁶ Folic acid intake during the first trimester is associated with reduced language delay, increased communication and verbal skills, and increased cognitive performance.^{55,57,58} Furthermore, supplementation with folic acid during the second and third trimesters decreased the levels of homocysteine in maternal and cord blood, although outcomes in offspring were not studied.⁸¹ In animal models, offspring from rodents deficient in methyl donors during pregnancy but supplemented with folic acid during late gestation had reduced risk for structural and functional defects in the CNS during the prenatal period.⁸² Additionally, reduced intake of folate in pregnant mice impaired short-term memory in offspring and increased apoptosis in the hippocampus.⁸³ Folic acid supplementation in late gestation restored expression of specific microRNAs in the brain in methyl donor-deficient animals.⁸² Expression of a number of

genes related to neuronal function is altered in the brain of offspring after folic acid supplementation.⁵⁰

In naturally occurring food sources, a combination of multiple methyl-donor nutrients is typically found within a single food. Additionally, methyl-donor nutrients work in concert with each other within 1-carbon metabolism. Therefore, supplementation with mixtures of methyl-donor nutrients, as opposed to supplementation with single nutrients, has been examined as well. Children of mothers supplemented with a mixture of micronutrients that included multiple methyl-donor nutrients had increased procedural memory and higher general intellectual ability at 10 years of age.⁶⁰ A study of maternal methyl-donor nutrient consumption found positive associations between folic acid intake and cognition in children at 3 years, but no associations were found for choline, betaine, or methionine consumption.⁵⁸ In rodents, methyl-donor supplementation during pregnancy and lactation increased DNA methylation in the brain^{42,84} and normalized the increased preference for palatable foods observed in offspring from high-fat diet-fed dams.⁴² Postnatal supplementation with multiple methyl-donor nutrients during lactation (postnatal day 2–9) restored methionine levels and reduced early-life-stress-induced cognitive impairments in adult animals.³⁹ Additionally, maternal micronutrient imbalance (excessive folic acid and/or vitamin B₁₂ deficiency) leads to global DNA hypomethylation and reduced expression of neurotrophins in the brain of offspring.^{14,85}

Early postnatal supplementation

Supplementation with methyl-donor nutrients during early life (infancy, childhood, or adolescence) is

Table 2 Clinical studies in humans linking methyl-donor supplementation with neurodevelopment and cognition

Reference	Behavioral test	Supplementation period	Nutrient(s) supplemented or analyzed	Outcomes
Boeke et al. (2013) ⁵³	Offspring visual memory and intelligence	Pregnancy (1st and 2nd trimesters)	Analyzed: choline, vitamin B ₁₂ , betaine, and folate (maternal intake questionnaire)	Increased gestational choline intake was associated with better visual memory at age 7; no associations for other methyl donors
Wu et al. (2012) ⁵¹	Bayley Scales of Infant Development, Third Edition	Pregnancy (2nd trimester)	Analyzed: choline, betaine, dimethylglycine, methionine, homocysteine, cysteine, vitamin B ₁₂ , holotranscobalamin, and folate (maternal plasma)	Maternal plasma free choline and betaine in the first half of pregnancy associated with increased infant cognitive test scores
Valera-Gran et al. (2014) ⁵²	Bayley Scales of Infant Development (1 y)	Pregnancy	Analyzed: folic acid (maternal intake questionnaire)	Children from mothers who consumed over 5000 µg of folic acid daily had lower psychomotor scores and increased risk for delayed psychomotor development
Signore et al. (2008) ⁵³	Intelligence quotient (5 y)	Pregnancy	Analyzed: choline (maternal cord blood)	No correlations found
Julvez et al. (2009) ⁵⁴	McCarthy Scale of Children's Abilities (4 y)	Pregnancy (1st trimester)	Analyzed: folic acid supplementation (maternal intake questionnaire)	Reported folic acid supplement use during pregnancy was associated with improved neurodevelopment and decreased inattention symptoms
Roth et al. (2011) ⁵⁵	Children's language competency (3 y)	Pregnancy (4 wk before conception to 8 wk after conception)	Analyzed: folic acid supplementation (maternal intake questionnaire)	Folic acid supplementation during early pregnancy was associated with reduced risk of severe language delay
Veena et al. (2010) ⁵⁶	Cognitive function assessment—memory, attention and fluid reasoning (9–10 y)	Pregnancy (30 wk)	Analyzed: folic acid, vitamin B ₁₂ and homocysteine (maternal plasma)	Increased folic acid concentrations associated with increased cognitive test scores; no associations were seen with vitamin B ₁₂ or homocysteine
Chatzi et al. (2012) ⁵⁷	Bayley Scales of Infant Development, Third Edition (18 mo)	Pregnancy (14–18 wk)	Analyzed: folic acid supplementation (maternal intake questionnaire and RBC concentration)	Folic acid supplementation (5 mg/d) was associated with enhanced vocabulary development, communication skills, and verbal comprehension
Villamor et al. (2012) ⁵⁸	Peabody Picture Vocabulary Test III and Wide Range Assessment of Visual Motor Abilities (3 y)	Pregnancy (1st and 2nd trimesters)	Analyzed: folic acid, vitamin B ₁₂ , choline, betaine, and methionine (maternal intake questionnaire)	Higher folate intake in 1st trimester was associated with higher receptive language; weak inverse association between vitamin B ₁₂ intake during 2nd trimester and receptive language; no association with choline, betaine, or methionine
Cheatham et al. (2012) ⁵⁹	Short-term visuospatial memory, long-term episodic memory, language development, and global development	Pregnancy + lactation (18 wk gestation to 90 d post partum)	Supplemented: phosphatidylcholine	No effects seen

(continued)

Table 2 Continued

Reference	Behavioral test	Supplementation period	Nutrient(s) supplemented or analyzed	Outcomes
Prado et al. (2017) ⁶⁰	General intellectual ability, declarative memory, procedural memory, executive function, academic achievement, fine motor dexterity, and socioemotional health (9–12 y)	Pregnancy + lactation (pregnancy + 3 mo post partum)	Supplemented: multiple micronutrients (iron, folic acid, retinol, vitamins D, E, C, B ₁ , B ₂ , B ₆ , B ₁₂ , zinc, copper, selenium, and iodine) or iron and folic acid	Supplementation with multiple micronutrients increased child cognitive development more than that with iron and folic acid alone
Ross et al. (2016) ⁶¹	Child Behavior Checklist (40 mo)	Pregnancy + lactation (2nd trimester to 3 mo)	Supplemented: phosphatidylcholine	Phosphatidylcholine supplementation decreased attention problems and social withdrawal
Nguyen et al. (2016) ⁶²	Memory, executive function, attention, and hyperactivity	Early life (5–10 y)	Supplemented: choline (6 wk)	No effects seen
Strain et al. (2013) ⁶³	Finger tapping test, Preschool Language Scale–Revised, Woodcock-Johnson Test of Scholastic Achievement, Child Behavior Checklist, and Kaufman Brief Intelligence Test	Early life (5 y)	Analyzed: choline, betaine, dimethylglycine, methionine, and homocysteine (plasma)	No associations found
Rauh-Pfeiffer et al. (2014) ⁶⁴	HAWIVA-III, WPPSI-III, Kaufman Assessment Battery for Children	Early life (4–6 y)	Supplemented: folic acid, riboflavin, pyridoxine, cobalamin, and calcium lactate pentahydrate (3 mo)	Supplementation decreased plasma homocysteine concentration, but no changes were seen in cognitive performance
Lefèvre-Arbogast et al. (2016) ³⁰	Dementia	Adult	Analyzed: folic acid, vitamins B ₆ and B ₁₂ (24-h recall)	Higher folate intakes associated with decreased risk of dementia; no associations found for vitamin B ₆ or B ₁₂
Morris et al. (2010) ⁶⁵	Wechsler Adult Intelligence Scale III	Adult (≥ 60 y)	Analyzed: folic acid, vitamin B ₁₂ , and 5-MeTHF (serum)	Normal vitamin B ₁₂ status together with higher serum 5MeTHF associated with higher cognitive test scores
Morris et al. (2007) ⁶⁶	Wechsler Adult Intelligence Scale III	Adult (≥ 60 y)	Analyzed: folic acid and vitamin B ₁₂ (serum)	Low vitamin B ₁₂ together with high serum folate associated with cognitive impairment; normal vitamin B ₁₂ status together with high serum folate associated with protection against cognitive impairment
Wald et al. (2010) ⁶⁷	Cognitive function test scores (3 y from start of treatment)	Adult (≥ 45 y)	Analyzed: meta-analysis of 9 placebo-controlled randomized trials, supplementing folic acid with or without other B vitamins	No associations found
Durga et al. (2007) ⁶⁸	Performance for memory, sensorimotor speed, complex speed, information processing speed, and word fluency	Adult (50–70 y)	Supplemented: folic acid (3 y)	3-y changes in memory, information processing speed, and sensorimotor speed were significantly better in the folic acid-supplemented group

(continued)

Table 2 Continued

Reference	Behavioral test	Supplementation period	Nutrient(s) supplemented or analyzed	Outcomes
Walker et al. (2012) ⁶⁹	Telephone Interview for Cognitive Status–Modified, the Brief Test of Adult Cognition by Telephone, and the Informant Questionnaire on Cognitive Decline in the Elderly	Adult (60–74 y)	Supplemented: folic acid and vitamin B ₁₂ (2 y)	Long-term supplementation of daily oral folic acid and vitamin B ₁₂ improved immediate and delayed memory performance; no differences seen in orientation, attention, semantic memory, processing speed, or informant reports
de Koning et al. (2016) ⁷⁰	Geriatric Depression Scale	Adult (≥ 65 y)	Supplemented: folic acid and vitamin B ₁₂ (2 y)	No changes in depressive symptoms observed
Zhang et al. (2017) ³¹	MMSE	Adult (≈ 75 y)	Analyzed: meta-analysis of 77 studies, supplementing folic acid, and vitamin B ₁₂ and/or vitamin B ₆ (supplementation length varied from 6 to 18 mo)	No associations found
Clarke et al. (2014) ⁷¹	Specific cognitive domains and MMSE	Adult ($\approx \geq 60$ y)	Analyzed: meta-analysis of 11 studies, supplementing B vitamins (supplementation length varied from 0.3 to 7 y)	No associations found
Porter et al. (2017) ⁷²	MMSE, RBANS, and the Frontal Assessment Battery	Adult (≥ 60 y)	Analyzed: folate, vitamins B ₁₂ , B ₆ , B ₂ , and homocysteine (at 5-y follow-up)	Lower vitamin B ₆ status was associated with accelerated rate of cognitive decline as assessed by RBANS and MMSE; lower vitamin B ₂ status was associated with accelerated rate of cognitive decline as assessed by RBANS, but not MMSE

Abbreviations: HAWIVA-III, Hannover-Wechsler-Intelligenztest für das Vorschulalter; MMSE, Mini Mental State Examination; RBC, red blood cell; RBANS, Repeatable Battery for the Assessment of Neuropsychological Status; WPPSI-III, Wechsler Preschool and Primary Scale of Intelligence, 3rd edition; 5-MeTHF, 5-methyltetrahydrofolate.

relatively understudied, as the majority of supplementation studies have focused on supplementation during pregnancy or later life. Infants supplemented with choline were shown to have fewer attention problems and less social withdrawal at 40 months.⁶¹ However, if administered at a later time point (to school-aged children), 6 weeks of choline supplementation did not improve cognition.⁶² Similarly, 3 months of mixed B vitamin supplementation in kindergarten children improved folate and homocysteine status but did not affect cognitive performance.⁶⁴ However, betaine status, but not choline status, in 5-year-olds was positively associated with total language scores.⁶³ In rodent models, rat pups deficient in folate from birth to 3 weeks of age exhibited deficits in long-term memory, spatial learning, and set-shifting.⁸⁶ Three weeks of 90% methyl-donor depletion increased depression-like behaviors in the forced swim test, while 3 weeks of methyl-donor supplementation increased DNA methylation in the amygdala and reduced

depression-like behaviors.⁸⁷ Similarly, 3 weeks of folic acid supplementation prevented hypoxia-ischemia-induced anxiogenic and memory impairments, possibly by partial recovery of Na⁺, K⁺-ATPase in the frontal cortex.⁴⁵ Finally, early-life postnatal supplementation with methyl-donor nutrients during weeks 3 to 6 of life in female mice increased motivation and learning in an operant chamber task.⁴⁴ Collectively, these studies demonstrate that methyl-donor supplementation can affect neurodevelopment and cognition beyond the well-established critical period of pregnancy; however, studies in humans indicate the window of opportunity for intervention may be reduced by the time children have reached school age.

Supplementation in adults

Memory loss, increased neurological disease risk, and increased oxidative stress in the brain are characteristic

of aging adults. In aged rats, 8 weeks of folic acid supplementation improved memory and decreased lipid peroxidation, a marker of aging, in multiple brain regions.⁴⁸ In adult rats, 7 days of folic acid treatment decreased locomotor activity and lipid peroxidation in the hippocampus in a pharmacological model of manic behavior.⁴⁹ Furthermore, 1 week of methyl-donor supplementation in mice increased motivation to respond in an operant task, although only if animals had been exposed to a high-fat diet during pregnancy/lactation or had been previously exposed to methyl-donor nutrients early in life.⁴⁴ Lastly, 18 weeks of dietary methyl-donor supplementation normalized depression-like behaviors induced by maternal separation during lactation.⁴⁷

While studies of methyl-donor supplementation in aged animals are limited, the use of methyl-donor nutrients in clinical trials is extensive. The results, however, are mixed. A meta-analysis examined whether 6 months of folic acid supplementation, with or without other B vitamins, altered cognitive function (ie, memory speed, language, and executive function) in healthy individuals and found no changes in cognition.⁶⁷ However, a study of 3 years of folic acid supplementation in healthy individuals reported higher scores for memory, information processing speed, and sensorimotor speed.⁶⁸ In a trial of 2-year supplementation with folic acid and B₁₂, improved cognitive functioning, including improved immediate and delayed memory performance, was observed.⁶⁹ However, in 2 separate trials, 2 years of B₁₂ and folic acid supplementation in older adults did not reduce depressive symptoms.^{70,88} A recent meta-analysis identified 77 human trials designed to examine whether B vitamin supplementation reduced homocysteine levels and improved cognition in elderly patients with Alzheimer disease or dementia.³¹ After filtering and selection, 4 randomized controlled trials were included in the analysis. With the Mini-Mental State Examination score used as a measure of cognition, the meta-analysis found that B vitamins do reduce homocysteine levels; however, there was no overall improvement in cognition in these patients.³¹ Similarly, another meta-analysis of 11 studies involving 22 000 study participants failed to find an effect of B vitamin supplementation on cognition, even though verified reductions in homocysteine and increases in folate were observed.⁷¹ Future longitudinal studies may focus on whether these B vitamins could perhaps slow the progression of cognitive decline or be efficacious in select clinical populations.⁸⁹ Additionally, a study showed that lower concentrations of vitamin B₆ and riboflavin (B₂) were associated with an accelerated rate of cognitive decline as assessed by the Repeatable Battery for the Assessment of Neuropsychological Status, but the same association with vitamin B₂ was not found

when the Mini-Mental State Examination was used to measure cognition.⁷²

Adverse effects of supplementation

The Swedish physician Paracelsus said, "The dose makes the poison," which is a basic principle of toxicology and applies equally to the study of dietary supplementation. Owing to both supplementation of processed food products and adherence to dietary recommendations for nutrient consumption, 5% of the US adult population and between 30% and 66% of children aged 1 to 13 years consume more than the recommended daily dose of methyl vitamins.^{90,91} As this subset of the population emerged, studies began showing the detrimental effects of excessive methyl-donor supplementation. The current recommendation for folic acid intake is 400 µg/d for adult males and nonpregnant females. The recommendation increases to 600 µg/d for pregnant females and 500 µg/d for lactating females. Offspring of women who consumed higher-than-recommended amounts of folic acid during pregnancy (5000 µg/d, almost 10 times the recommended daily allowance) had lower psychomotor scores at 1 year of age.⁵² In mice, offspring of dams fed high levels of folic acid (10 times the control, 20 mg/kg of diet) during gestation displayed altered development of the cortical layers at E17.5 and short-term memory impairment and decreased hippocampal size at 3 weeks.⁴³ High-vitamin diets during pregnancy have been shown to increase the risk of metabolic syndrome in offspring by altering the neural pathways involved in energy homeostasis and feeding,^{92,93} but this could be prevented by providing a postweaning diet high in multivitamins or folic acid.⁹⁴ Excessive folic acid intake (4 times the control, 8.0 mg/kg of diet) during adolescence has been correlated with deficits in motivation and spatial memory tasks.⁴⁶ Another factor that affects response to methyl-donor supplementation is genetic variations in the enzymes that metabolize the nutrients, such as methylenetetrahydrofolate reductase (MTHFR). In dams with a MTHFR deficiency, methyl-donor supplementation was associated with embryonic delay and growth retardation at E10.5 in their offspring, but longer-term outcomes were not studied.⁹⁵ It is evident that a genetic and environmental interaction exists in which concentrations of methyl-donor nutrients are no longer healthful, but are harmful. Studying these interactions remains critical (reviewed by Shorter et al⁹⁶).

CONSIDERATIONS FOR FUTURE RESEARCH

It is clear that both excessive and inadequate levels of methyl levels are detrimental to neurodevelopment and

cognition. Further research is needed to understand and standardize which nutrients to provide supplementally, which concentrations of each nutrient are appropriate, how long a supplement should be taken, and which populations should receive supplementation as well as to determine whether methyl-donor supplementation can alter CNS development and long-term cognition. Many of the basic research studies point to methyl-donor supplementation as a way to modulate cognition, but the current clinical literature is inconclusive.⁹⁷ A number of experimental variables need to be thoughtfully considered to ensure the robustness of research in this field. The focus here is on animal research, but similar considerations are applicable to human clinical research as well.

Single-nutrient vs mixed-nutrient supplementation

In reviewing the current supplements used to enhance brain health, many studies promote single-nutrient supplementation while others tout a mixed-nutrient approach. The mixed-nutrient approach may be more relevant to what is found in naturally occurring food sources (which contain multiple methyl donors). Additionally, there is the unique relationship between folate deficiency and vitamin B₁₂ deficiency, wherein the surplus of one may mask a deficiency of the other (ie, excess folate/folic acid may mask a vitamin B₁₂ deficiency). Generally, cellular folate is present in low concentrations; therefore, regenerating tetrahydrofolate from 5-methyltetrahydrofolate is extremely important for 1-carbon metabolism and regulation of cellular homocysteine levels. This regeneration can occur in 2 ways. One is dependent on the availability of vitamin B₁₂ to act as a coenzyme for methionine synthase. When vitamin B₁₂ is not present, the cell has increased concentrations of both 5-methyltetrahydrofolate, which has been shown to impair DNA and RNA synthesis,⁹⁸ and homocysteine, high levels of which have been observed in many neurological disorders.^{5,24,35,99} Folic acid supplementation has been shown to decrease the homocysteine concentration, the biomarker routinely used to determine cellular levels of vitamin B₁₂ and folic acid. However, when high folate levels exist together with low vitamin B₁₂ levels, individuals present with anemia, macrocytosis, and cognitive impairment.^{65,66,100} Therefore, a mixed-nutrient approach to methyl-donor supplementation may mitigate these masking effects.

One-carbon metabolism and the brain

The liver is responsible for the vast majority of metabolic processes in the body. Therefore, 1-carbon

metabolism has largely been functionally characterized within the liver. It is widely thought that these methyl-donor nutrients are metabolized by the liver, and then metabolites are transported by the blood to be distributed throughout the body, passing through the blood-brain barrier to reach the brain. However, because enzymes that mediate 1-carbon metabolism are present within the brain, this would suggest that local 1-carbon metabolism can also occur within the brain.

Even though 1-carbon metabolism in the liver is well studied and research to date has provided an understanding of how perturbations such as diet or disease alter this metabolism, 1-carbon metabolism in the brain has not been well characterized. It is thought, on the basis of increased homocysteine concentrations in plasma and cerebrospinal fluid, that 1-carbon metabolism in the brain is disrupted in various neurological diseases such as Alzheimer disease, depression, and schizophrenia.^{101–104} This relies, however, on the assumption that peripheral metabolite levels are reliable biomarkers of brain levels, which has not yet been demonstrated. Importantly, a few functional differences between 1-carbon metabolism in the brain and 1-carbon metabolism in the liver have been found. As recently as 2006, there remained controversy about whether the brain contained a functional transsulfuration pathway, important for the elimination of reactive oxygen species.²⁶ Because the brain consumes a relatively high level of oxygen, effective strategies for the elimination of high levels of reactive oxygen species are necessary. Possible impairments to the brain transsulfuration pathway are found both in patients with Alzheimer disease and in those with Parkinson disease.²⁶ Furthermore, levels of cystathionine are higher in the brain than in other organs, while γ -cystathionase activity in the brain is 100-fold lower than that in the liver. It is now known that the brain contains a functional transsulfuration pathway, but these 2 functional differences provide evidence that this pathway may function differently in the brain than in the liver.

Additionally, the brain lacks the enzyme betaine-homocysteine methyltransferase.²⁴ Therefore, it lacks the ability to utilize betaine to resynthesize methionine from homocysteine, a pathway present in the liver. This may indicate that the brain, compared with the liver, has increased reliance on the presence of vitamin B₁₂ to aid in the maintenance of cellular methionine levels. There is evidence that the brain is extremely resilient when nutrient deficiencies are present. One study found that, when rats were fed a folate-free diet, total folate concentrations in the liver and kidney decreased by 60%, but levels of folate in the brain remained the same throughout the study.¹⁰⁵ Additionally, a recent study showed that, when folate deficiency was induced, the

brain—as compared with plasma, erythrocytes, liver, kidney, and spleen—maintained folate metabolite levels better than the other organs.¹⁰⁶ These studies indicate a possible maintenance system or protective mechanism present in the brain but not in other organs. One potential mechanism could involve folate-binding proteins, which are expressed at high levels within the choroid plexus^{107,108} and may contribute to higher folate levels in the brain at the expense of levels in other tissues. This same phenomenon may also occur in other organs with high expression of folate-binding proteins, such as the placenta,^{109,110} which would result in folate being provided to the offspring at the expense of the mother. However, it has also been shown that vitamin B₁₂ levels in the brain decreased during dietary folate deficiency.¹¹¹ Current gaps in the literature include a lack of studies evaluating brain levels of 1-carbon metabolites in response to dietary changes. There is also a need to determine whether plasma levels of metabolites reflect changes in brain levels, since findings of an association could support the potential use of plasma levels as a biomarker for brain dysfunction.

It is well known that the brain is composed of multiple cell types, including neurons, astrocytes, microglia, and endothelial cells, that work coordinately. Functional differences in various cell types could alter 1-carbon metabolism.⁵ For example, the reliance on glucose to provide serine for 1-carbon metabolism is higher in the brain than in other organs, since serine does not easily pass through the blood–brain barrier.⁵ Additionally, the lack of betaine-homocysteine methyltransferase in the brain alters 1-carbon metabolism by eliminating the ability to utilize betaine directly to remethylate homocysteine. Neurons, glia, and astrocytes could also conceivably utilize the same nutrients in different ways because of their functional differences. One possible example is the increased reliance of the brain on astrocytes and oligodendrocytes to mediate transsulfuration pathway reactions. The brain consumes a disproportionate amount of the body's oxygen and therefore must eliminate copious amounts of reactive oxygen species. Synthesis of glutathione is critical to this elimination. In turn, glutathione synthesis is dependent on the cysteine concentration. Excitatory amino acid transporter 3, the main transporter of extracellular cysteine, has been found in greater densities on astrocytes and oligodendrocytes,¹¹² possibly indicating increased participation of these 2 cell types in transsulfuration pathway-mediated elimination of reactive oxygen species. Further characterization of the roles each cell type plays in brain 1-carbon metabolism will be important in developing a full appreciation of the role of 1-carbon metabolism in the brain.

Influence of sex on outcomes

The pervasiveness of sex differences in the brain and neurological disorders warrants a discussion in this context. While investigations of sex differences are increasingly included in preclinical research and are typically included in the human clinical literature, analysis of both sexes remains underrepresented in the current animal literature. Only 4 of the 14 studies shown in Table 1 included a comparison between sexes, while 8 focused on a single sex, 1 studied combined sexes without analysis, and another did not report the sex of the animals. Sex differences within 1-carbon metabolism have not been extensively examined, although some reports suggest that sex is an important variable with regard to the study of 1-carbon metabolism and brain function. Sex was found to influence vitamin B₁₂ and 1-carbon-related enzymes as well as plasma homocysteine levels in the fetal liver, and sex moderated the effect of maternal smoking on these outcomes.¹¹³ Plasma levels of homocysteine were found to correlate with some brain metabolite levels (as determined through imaging) in elderly females, but not males.¹¹⁴ Furthermore, in an animal study involving supplementation of high-dose folic acid to pregnant dams, the expression of transcription factors related to neuronal function in whole brain samples was oppositely affected in males (decreased) compared with females (increased).¹¹⁵ In addition, in a mouse model with a deletion of the *Mthfr* gene, early-life manipulations, either neonatal stress¹¹⁶ or administration of a γ -aminobutyric acid-potentiating drug,¹¹⁷ were found to have sex-specific effects on behavior.

Sex differences in DNA methylation patterns in the brain have also been noted. At postnatal day 1, female rats were found to have significantly higher activity of DNA methyltransferase 1 as well as an increase in DNA methylation¹¹⁸ in the preoptic area of the hypothalamus. Sex differences in the expression of genes that either increase DNA methyltransferases¹¹⁹ or decrease (Gadd45b) DNA methylation¹²⁰ have been demonstrated in the amygdala, and sex differences in promoter-specific DNA methylation of the *bdnf* gene have been observed within the prefrontal cortex.¹²¹ It has been well characterized that sex alters the prevalence of many neurological disorders, including Parkinson disease, Alzheimer disease, cognitive decline, autism spectrum disorders, mood and anxiety disorders, major depression, trauma-related disorders, depressive disorders, autoimmune disease affecting the nervous system, attention-deficit/hyperactivity disorder, and neurodegenerative disorders.¹²² Collectively, these findings suggest that understanding sex differences in 1-carbon metabolism in the brain, both at baseline and

in response to methyl-donor supplementation, may increase the current understanding of sex differences in neurodevelopmental outcomes.

In mammals, the presence of *SRY* on the male Y chromosome drives the development of the testis. Gonadal sex hormones (androgens, estrogens, and progestins) then drive the sexual differentiation. During development, the male brain is exposed to a testosterone surge, which is converted to estradiol. At the same time, in females, the estradiol levels remain low.¹²³ Within the brain, these sex hormones are known to influence neuroanatomy, neurochemistry, and neuron structure.¹²² One interesting difference between the sexes is the growth speed of the cortex. During development, cortical volume increases faster in males than in females.¹²⁴ This growth difference is hypothesized to drive, in part, the predisposition in males for autism spectrum disorders.¹²⁵ Additionally, during development, maternal care, estrogen receptor expression, and neonatal hormone exposure all have been shown to correlate with sex-specific differences in DNA methylation, which in turn can influence development and behavior.¹²⁶

Sex hormones can also have effects during adulthood, when females are exposed to high levels of estradiol and progesterone and males are exposed to high levels of testosterone. To add complexity, the female estrus cycle alters the levels of the female sex hormones, which is known to change behavior.^{127–131} Additionally, steroid hormone receptors, which can be found in different densities throughout the brain, are known to act as transcription factors and influence epigenetic machinery (coactivators and corepressors).^{123,126} Interestingly, there is even a sex-specific difference in the expression of these coactivators.¹³² Monitoring the estrus cycle would inform whether observed sex differences remain throughout the entire estrus cycle or are stronger at one point versus another, while testing at a standard stage of the estrus cycle may decrease the variance that could be observed in female study participants. Lastly, the presence of sex chromosomes, or genetic sex (XX vs XY), can also influence neurodevelopment and sexual differentiation in the brain (reviewed by McCarthy and Arnold¹³³). Thus, genetic sex and the presence of sex hormones during development and throughout adulthood both have the potential to influence the physiological endpoints of interest. The inclusion and analysis of both sexes will prove essential to understanding how supplementation and dietary interventions alter neurodevelopment and cognition.

CONCLUSION

The current literature suggests that dietary methyl-donor supplementation is a promising therapeutic

strategy with the potential to improve or protect cognition and neurodevelopment in vulnerable populations. Earlier diagnosis of nutrient deficiencies and excesses, understanding critical periods for intervention, and identification of which populations would most benefit from supplementation could all help to increase the efficacy of supplementation. Ensuring robustness in animal studies through the careful consideration of diet composition, tissue and cellular specificity, and sex differences will significantly advance the understanding of how methyl-donor supplementation affects neurodevelopment on a mechanistic level as well as increase the potential for translation to at-risk human populations.

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REFERENCES

1. Moody L, Chen H, Pan Y-X. Early-life nutritional programming of cognition—the fundamental role of epigenetic mechanisms in mediating the relation between early-life environment and learning and memory process. *Adv Nutr*. 2017;8:337–350.
2. Chen Y, Baram TZ. Toward understanding how early-life stress reprograms cognitive and emotional brain networks. *Neuropsychopharmacology*. 2016;41:197–206.
3. Li E. Chromatin modification and epigenetic reprogramming in mammalian development. *Nat Rev Genet*. 2002;3:662–673.
4. Fox JT, Stover PJ. Folate-mediated one-carbon metabolism. *Vitam Horm*. 2008;79:1–44.
5. Ducker GS, Rabinowitz JD. One-carbon metabolism in health and disease. *Cell Metab*. 2016;25:27–42.
6. Tibbetts AS, Appling DR. Compartmentalization of mammalian folate-mediated one-carbon metabolism. *Annu Rev Nutr*. 2010;30:57–81.
7. Cohen EL, Wurtman RJ. Brain acetylcholine: control by dietary choline. *Science*. 1976;191:561–562.
8. Klinkenberg I, Sambeth A, Blokland A. Acetylcholine and attention. *Behav Brain Res*. 2011;221:430–442.
9. Savelkoul PJM, Janickova H, Kuipers AAM, et al. A specific multi-nutrient formulation enhances M1 muscarinic acetylcholine receptor responses in vitro. *J Neurochem*. 2012;120:631–640.
10. Zeisel SH, Niculescu MD. Perinatal choline influences brain structure and function. *Nutr Rev*. 2006;64:197–203.
11. Scott JM. Folate-vitamin B₁₂ interrelationships in the central nervous system. *Proc Nutr Soc*. 1992;51:219–224.
12. Anderson OS, Sant KE, Dolinoy DC. Nutrition and epigenetics: an interplay of dietary methyl donors, one-carbon metabolism and DNA methylation. *J Nutr Biochem*. 2012;23:853–859.
13. Niculescu MD, Zeisel SH. Diet, methyl donors and DNA methylation: interactions between dietary folate, methionine and choline. *J Nutr*. 2002;132(8 suppl):2335S–2335S.
14. Sable P, Kale A, Joshi A, et al. Maternal micronutrient imbalance alters gene expression of BDNF, NGF, TrkB and CREB in the offspring brain at an adult age. *Int J Dev Neurosci*. 2014;34:24–32.

15. Fernàndez-Roig S, Lai S-C, Murphy MM, et al. Vitamin B₁₂ deficiency in the brain leads to DNA hypomethylation in the TCbIR/CD320 knockout mouse. *Nutr Metab (Lond)*. 2012;9:41. doi:10.1186/1743-7075-9-41
16. Ly A, Hoyt L, Crowell J, et al. Folate and DNA methylation. *Antioxid Redox Signal*. 2012;17:302–326.
17. Niculescu MD, Craciunescu CN, Zeisel SH. Dietary choline deficiency alters global and gene-specific DNA methylation in the developing hippocampus of mouse fetal brains. *FASEB J*. 2006;20:43–49.
18. Tomizawa H, Matsuzawa D, Ishii D, et al. Methyl-donor deficiency in adolescence affects memory and epigenetic status in the mouse hippocampus. *Genes Brain Behav*. 2015;14:301–309.
19. Breimer LH, Nilsson TK. Has folate a role in the developing nervous system after birth and not just during embryogenesis and gestation?. *Scand J Clin Lab Invest*. 2012;72:185–191.
20. Araújo JR, Martel F, Borges N, et al. Folate and aging: role in mild cognitive impairment, dementia and depression. *Ageing Res Rev*. 2015;22:9–19.
21. Zhang Y, Hodgson NW, Trivedi MS, et al. Decreased brain levels of vitamin B12 in aging, autism and schizophrenia. *PLoS One*. 2016;11:e0146797. doi:10.1371/journal.pone.0146797
22. Grayson DR, Guidotti A. The dynamics of DNA methylation in schizophrenia and related psychiatric disorders. *Neuropsychopharmacology*. 2012;38:138–166.
23. Boeke CE, Gillman MW, Hughes MD, et al. Choline intake during pregnancy and child cognition at age 7 years. *Am J Epidemiol*. 2013;177:1338–1347.
24. Bottiglieri T. Homocysteine and folate metabolism in depression. *Prog Neuropsychopharmacol Biol Psychiatry*. 2005;29:1103–1112.
25. Suh JR, Herbig AK, Stover PJ. New perspectives on folate catabolism. *Annu Rev Nutr*. 2001;21:255–282.
26. Vitvitsky V, Thomas M, Ghorpade A, et al. A functional transsulfuration pathway in the brain links to glutathione homeostasis. *J Biol Chem*. 2006;281:35785–35793.
27. Jiang B, Yao G, Ding C, et al. Serum homocysteine, folic acid and VitB₁₂ in patients with mild cognitive impairment. 2017;10:3731–3736.
28. Setién-Suero E, Suárez-Pinilla M, Suárez-Pinilla P, et al. Homocysteine and cognition: a systematic review of 111 studies. *Neurosci Biobehav Rev*. 2016;69:280–298.
29. Kóbe T, Witte AV, Schnelle A, et al. Vitamin B-12 concentration, memory performance, and hippocampal structure in patients with mild cognitive impairment. *Am J Clin Nutr*. 2016;103:1045–1054.
30. Lefèvre-Arbogast S, Féart C, Dartigues J-F, et al. Dietary B vitamins and a 10-year risk of dementia in older persons. *Nutrients*. 2016;8:761. doi:10.3390/nu8120761
31. Zhang D-M, Ye J-X, Mu J-S, et al. Efficacy of vitamin B supplementation on cognition in elderly patients with cognitive-related diseases. *J Geriatr Psychiatry Neurol*. 2017;30:50–59.
32. Blom HJ. Folic acid, methylation and neural tube closure in humans. *Birth Defects Res Part A Clin Mol Teratol*. 2009;85:295–302.
33. Zeisel SH. Importance of methyl donors during reproduction. *Am J Clin Nutr*. 2009;89:673S–677S.
34. McGarel C, Pentieva K, Strain JJ, et al. Emerging roles for folate and related B-vitamins in brain health across the lifecycle. *Proc Nutr Soc*. 2015;74:46–55.
35. Kennedy DO. B vitamins and the brain: mechanisms, dose and efficacy—a review. *Nutrients*. 2016;8. doi:10.3390/nu8020068
36. Morris MS. The role of B vitamins in preventing and treating cognitive impairment and decline. *Adv Nutr*. 2012;3:801–812.
37. Thomas JD, Abou EJ, Dominguez HD. Prenatal choline supplementation mitigates the adverse effects of prenatal alcohol exposure on development in rats. *Neurotoxicol Teratol*. 2009;31:303–311.
38. Thomas JD, Idris NM, Monk BR, et al. Prenatal choline supplementation mitigates behavioral alterations associated with prenatal alcohol exposure in rats. *Birth Defects Res Part A Clin Mol Teratol*. 2010;88:827–837.
39. Naninck EFG, Oosterink JE, Yam K-Y, et al. Early micronutrient supplementation protects against early stress-induced cognitive impairments. *FASEB J*. 2016;9:fj.201600834R. doi:10.1096/fj.201600834R
40. Zhu CH, Wu T, Jin Y, et al. Prenatal choline supplementation attenuates spatial learning deficits of offspring rats exposed to low-protein diet during fetal period. *J Nutr Biochem*. 2016;32:163–170.
41. Langley EA, Krykbaeva M, Blusztajn JK, et al. High maternal choline consumption during pregnancy and nursing alleviates deficits in social interaction and improves anxiety-like behaviors in the BTBR T+^{itpr3f}/J mouse model of autism. *Behav Brain Res*. 2015;278:210–220.
42. Carlin J, George R, Reyes TM. Methyl donor supplementation blocks the adverse effects of maternal high fat diet on offspring physiology. *PLoS One*. 2013;8:e63549. doi:10.1371/journal.pone.0063549
43. Bahous RH, Jadavji NM, Deng L, et al. High dietary folate in pregnant mice leads to pseudo-MTHFR deficiency and altered methyl metabolism, with embryonic growth delay and short-term memory impairment in offspring. *Hum Mol Genet*. 2017;26:888–900.
44. McKee SE, Grissom NM, Herdt CT, et al. Methyl donor supplementation alters cognitive performance and motivation in female offspring from high-fat diet-fed dams. *FASEB J*. 2017;31:2352–2363.
45. Carletti JV, Deniz BF, Miguel PM, et al. Folic acid prevents behavioral impairment and Na⁺, K⁺-ATPase inhibition caused by neonatal hypoxia-ischemia. *Neurochem Res*. 2012;37:1624–1630.
46. Sittig LJ, Herzog LBK, Xie H, et al. Excess folate during adolescence suppresses thyroid function with permanent deficits in motivation and spatial memory. *Genes Brain Behav*. 2012;11:193–200.
47. Paternain L, Martisova E, Campión J, et al. Methyl donor supplementation in rats reverses the deleterious effect of maternal separation on depression-like behaviour. *Behav Brain Res*. 2016;299:51–58.
48. Singh R, Kanwar SS, Sood PK, et al. Beneficial effects of folic acid on enhancement of memory and antioxidant status in aged rat brain. *Cell Mol Neurobiol*. 2011;31:83–91.
49. Brocardo PS, Budni J, Pavesi E, et al. Folic acid administration prevents ouabain-induced hyperlocomotion and alterations in oxidative stress markers in the rat brain. *Bipolar Disord*. 2010;12:414–424.
50. Barua S, Chadman KK, Kuizon S, et al. Increasing maternal or post-weaning folic acid alters gene expression and moderately changes behavior in the offspring. *PLoS One*. 2014;9:e0101674. doi:10.1371/journal.pone.0101674
51. Wu BTF, Dyer RA, King DJ, et al. Early second trimester maternal plasma choline and betaine are related to measures of early cognitive development in term infants. *PLoS One*. 2012;7:1–8.
52. Valera-Gran D, García de la Hera M, Navarrete-Munoz EM, et al. Folic acid supplements during pregnancy and child psychomotor development after the first year of life. *JAMA Pediatr*. 2014;168:e142611. doi:10.1001/jamapediatrics.2014.2611
53. Signore C, Ueland PM, Troendle J, et al. Choline concentrations in human maternal and cord blood and intelligence at 5 y of age. *Am J Clin Nutr*. 2008;87:896–902.
54. Julvez J, Fortuny J, Mendez M, et al. Maternal use of folic acid supplements during pregnancy and four-year-old neurodevelopment in a population-based birth cohort. *Paediatr Perinat Epidemiol*. 2009;23:199–206.
55. Roth C, Magnus P, Schjølberg S, et al. Folic acid supplements in pregnancy and severe language delay in children. *JAMA*. 2011;306:1566–1573.
56. Veena SR, Krishnaveni GV, Srinivasan K, et al. Higher maternal plasma folate but not vitamin B-12 concentrations during pregnancy are associated with better cognitive function scores in 9- to 10-year-old children in South India. *J Nutr*. 2010;140:1014–1022.
57. Chatzi L, Papadopoulou E, Koutra K, et al. Effect of high doses of folic acid supplementation in early pregnancy on child neurodevelopment at 18 months of age: the mother-child cohort “Rhea” study in Crete, Greece. *Public Health Nutr*. 2012;15:1728–1736.
58. Villamor E, Rifas-Shiman SL, Gillman MW, et al. Maternal intake of methyl-donor nutrients and child cognition at 3 years of age. *Paediatr Perinat Epidemiol*. 2012;26:328–335.
59. Cheatham CL, Goldman BD, Fischer LM, et al. Phosphatidylcholine supplementation in pregnant women consuming moderate-choline diets does not enhance infant cognitive function: a randomized, double-blind, placebo-controlled trial. *Am J Clin Nutr*. 2012;96:1465–1472.
60. Prado EL, Sebayang SK, Apriatni M, et al. Maternal multiple micronutrient supplementation and other biomedical and socioenvironmental influences on children’s cognition at age 9–12 years in Indonesia: follow-up of the SUMMIT randomised trial. *Lancet Glob Health*. 2017;5:e217–e228.
61. Ross RG, Hunter SK, Hoffman MC, et al. Perinatal phosphatidylcholine supplementation and early childhood behavior problems: evidence for *CHRNA7* moderation. *Am J Psychiatry*. 2016;173:509–516.
62. Nguyen TT, Risbud RD, Mattson SN, et al. Randomized, double-blind, placebo-controlled clinical trial of choline supplementation in school-aged children with fetal alcohol spectrum disorders. *Am J Clin Nutr*. 2016;104:1683–1692.
63. Strain JJ, McSorley EM, van Wijngaarden E, et al. Choline status and neurodevelopmental outcomes at 5 years of age in the Seychelles Child Development Nutrition Study. *Br J Nutr*. 2013;110:330–336.
64. Rauh-Pfeiffer A, Handel U, Demmelmaier H, et al. Three-month B vitamin supplementation in pre-school children affects folate status and homocysteine, but not cognitive performance. *Eur J Nutr*. 2014;53:1445–1456.
65. Morris MS, Jacques PF, Rosenberg IH, et al. Circulating unmetabolized folic acid and 5-methyltetrahydrofolate in relation to anemia, macrocytosis, and cognitive test performance in American seniors. *Am J Clin Nutr*. 2010;91:1733–1744.
66. Morris MS, Jacques PF, Rosenberg IH, et al. Folate and vitamin B-12 status in relation to anemia, macrocytosis, and cognitive impairment in older Americans in the age of folic acid fortification. *Am J Clin Nutr*. 2007;85:193–200.
67. Wald DS, Kasteurirane A, Simmonds M. Effect of folic acid, with or without other B vitamins, on cognitive decline: meta-analysis of randomized trials. *Am J Med*. 2010;123:522e2–527e2.
68. Durga J, van Boxtel MPJ, Schouten EG, et al. Effect of 3-year folic acid supplementation on cognitive function in older adults in the FACIT trial: a randomised, double blind, controlled trial. *Lancet*. 2007;369:208–216.
69. Walker JG, Batterham PJ, Mackinnon AJ, et al. Oral folic acid and vitamin B-12 supplementation to prevent cognitive decline in community-dwelling older adults with depressive symptoms—the Beyond Ageing Project: a randomized controlled trial. *Am J Clin Nutr*. 2012;95:194–203.

70. de Koning E, van der Zwaluw N, van Wijngaarden J, et al. Effects of two-year vitamin B₁₂ and folic acid supplementation on depressive symptoms and quality of life in older adults with elevated homocysteine concentrations: additional results from the B-PROOF Study, an RCT. *Nutrients*. 2016;8:748. doi:10.3390/nu8110748
71. Clarke R, Bennett D, Parish S, et al. Effects of homocysteine lowering with B vitamins on cognitive aging: meta-analysis of 11 trials with cognitive data on 22,000 individuals. *Am J Clin Nutr*. 2014;100:657–666.
72. Porter K, Hughes CF, Hoey L, et al. Biomarkers of folate and related B-vitamins as predictors of cognitive decline in older Irish adults over a 5 year follow up period. *Proc Nutr Soc*. 2017;76:E4. doi:10.1017/S0029665117000040
73. Ladipo OA. Nutrition in pregnancy: mineral and vitamin supplements. *Am J Clin Nutr*. 2000;72(1 suppl):280S–290S.
74. McCann JC, Ames BN. An overview of evidence for a causal relation between iron deficiency during development and deficits in cognitive or behavioral function. *Am J Clin Nutr*. 2007;85:931–945.
75. Sandstrom NJ, Loy R, Williams CL. Prenatal choline supplementation increases NGF levels in the hippocampus and frontal cortex of young and adult rats. *Brain Res*. 2002;947:9–16.
76. Davison JM, Mellott TJ, Kovacheva VP, et al. Gestational choline supply regulates methylation of histone H3, expression of histone methyltransferases G9a (Kmt1c) and Suv39h1 (Kmt1a), and DNA methylation of their genes in rat fetal liver and brain. *J Biol Chem*. 2009;284:1982–1989.
77. Craciunescu CN, Johnson AR, Zeisel SH. Dietary choline reverses some, but not all, effects of folate deficiency on neurogenesis and apoptosis in fetal mouse brain. *J Nutr*. 2010;140:1162–1166.
78. Tran X, Kennedy BC, Pisansky MT, et al. Prenatal choline supplementation diminishes early-life iron deficiency-induced reprogramming of molecular networks associated with behavioral abnormalities in the adult rat hippocampus. *J Nutr*. 2016;146:484–493.
79. Kelley CM, Powers BE, Velazquez R, et al. Maternal choline supplementation differentially alters the basal forebrain cholinergic system of young-adult Ts65Dn and disomic mice. *J Comp Neurol*. 2014;522:1390–1410.
80. Jiang X, West AA, Caudill MA. Maternal choline supplementation: a nutritional approach for improving offspring health? *Trends Endocrinol Metab*. 2014;25:263–273.
81. McNulty B, McNulty H, Marshall B, et al. Impact of continuing folic acid after the first trimester of pregnancy: findings of a randomized trial of folic acid supplementation in the second and third trimesters. *Am J Clin Nutr*. 2013;98:92–98.
82. Geoffroy A, Kerek R, Pourié G, et al. Late maternal folate supplementation rescues from methyl donor deficiency-associated brain defects by restoring Let-7 and miR-34 pathways. *Mol Neurobiol*. 2017;54:5017–5033.
83. Jadavji NM, Deng L, Malysheva O, et al. MTHFR deficiency or reduced intake of folate or choline in pregnant mice results in impaired short-term memory and increased apoptosis in hippocampus of wild-type offspring. *Neuroscience*. 2015;300:1–9.
84. Cooney CA, Dave AA, Wolff GL. Maternal methyl supplements in mice affect epigenetic variation and DNA methylation of offspring. *J Nutr*. 2002;132(8 suppl):2393S–2400S.
85. Sable P, Randhir K, Kale A, et al. Maternal micronutrients and brain global methylation patterns in the offspring. *Nutr Neurosci*. 2015;18:30–36.
86. Berrocal-Zaragoza MI, Sequeira JM, Murphy MM, et al. Folate deficiency in rat pups during weaning causes learning and memory deficits. *Br J Nutr*. 2014;112:1323–1332.
87. McCoy CR, Jackson NL, Day J, et al. Genetic predisposition to high anxiety- and depression-like behavior coincides with diminished DNA methylation in the adult rat amygdala. *Behav Brain Res*. 2017;320:165–178.
88. Walker JG, Mackinnon AJ, Batterham P, et al. Mental health literacy, folic acid and vitamin B₁₂, and physical activity for the prevention of depression in older adults: randomised controlled trial. *Br J Psychiatry*. 2010;197:45–54.
89. McCaddon A, Miller JW. Assessing the association between homocysteine and cognition: reflections on Bradford Hill, meta-analyses, and causality. *Nutr Rev*. 2015;73:723–735.
90. Bailey RL, Dodd KW, Gahche JJ, et al. Total folate and folic acid intake from foods and dietary supplements in the United States: 2003–2006. *Am J Clin Nutr*. 2010;91:231–237.
91. Bailey RL, McDowell MA, Dodd KW, et al. Total folate and folic acid intakes from foods and dietary supplements of US children aged 1–13 y. *Am J Clin Nutr*. 2010;92:353–358.
92. Cho CE, Sánchez-Hernández D, Reza-López SA, et al. High folate gestational and post-weaning diets alter hypothalamic feeding pathways by DNA methylation in Wistar rat offspring. *Epigenetics*. 2013;8:710–719.
93. Pannia E, Cho CE, Kubant R, et al. A high multivitamin diet fed to Wistar rat dams during pregnancy increases maternal weight gain later in life and alters homeostatic, hedonic and peripheral regulatory systems of energy balance. *Behav Brain Res*. 2014;278C:1–11.
94. Cho CE, Sánchez-Hernández D, Reza-López SA, et al. Obesogenic phenotype of offspring of dams fed a high multivitamin diet is prevented by a post-weaning high multivitamin or high folate diet. *Int J Obes (Lond)*. 2013;37:1177–1182.
95. Pickell L, Brown K, Li D, et al. High intake of folic acid disrupts embryonic development in mice. *Birth Defects Res Part A Clin Mol Teratol*. 2011;91:8–19.
96. Shorter KR, Felder MR, Vrana PB. Consequences of dietary methyl donor supplements: is more always better? *Prog Biophys Mol Biol*. 2015;118:14–20.
97. Veena SR, Gale CR, Krishnaveni GV, et al. Association between maternal nutritional status in pregnancy and offspring cognitive function during childhood and adolescence: a systematic review. *BMC Pregnancy Childbirth*. 2016;16:220. doi:10.1186/s12884-016-1011-z
98. Herbert V, Zalusky R. Interrelations of vitamin B₁₂ and folic acid metabolism: folic acid clearance studies. *J Clin Invest*. 1962;41:1263–1276.
99. Mattson MP, Shea TB. Folate and homocysteine metabolism in neural plasticity and neurodegenerative disorders. *Trends Neurosci*. 2003;26:137–146.
100. Selhub J, Paul L. Folic acid fortification: why not vitamin B12 also? *Biofactors*. 2011;37:269–271.
101. Dayon L, Guiraud SP, Corthésy J, et al. One-carbon metabolism, cognitive impairment and CSF measures of Alzheimer pathology: homocysteine and beyond. *Alzheimers Res Ther*. 2017;9:43. doi:10.1186/s13195-017-0270-x
102. Smythies J. The role of abnormalities related to the one carbon cycle in depression and schizophrenia. *Neurosci Med*. 2012;3:101–106.
103. Gottfries J, Blennow K, Rychlik M. Characterization and interrelations of one-carbon metabolites in tissues, erythrocytes, and plasma in mice with dietary induced folate deficiency. *Nutrients*. 2017;9:462. doi:10.3390/nu9050462
104. Coppède F. One-carbon metabolism and Alzheimer's disease: focus on epigenetics. *Curr Genomics*. 2010;11:246–260.
105. Varela-Moreiras G, Selhub J. Long-term folate deficiency alters folate content and distribution differentially in rat tissues. *J Nutr*. 1992;122:986–991.
106. Kopp M, Morisset R, Rychlik M. Characterization and interrelations of one-carbon metabolites in tissues, erythrocytes, and plasma in mice with dietary induced folate deficiency. *Nutrients*. 2017;9:462. doi:10.3390/nu9050462
107. Wollack JB, Makori B, Ahlawat S, et al. Characterization of folate uptake by choroid plexus epithelial cells in a rat primary culture model. *J Neurochem*. 2008;104:1494–1503.
108. Holm J, Hansen SI, Hoier-Madsen M, et al. High-affinity folate binding in human choroid plexus. Characterization of radioligand binding, immunoreactivity, molecular heterogeneity and hydrophobic domain of the binding protein. *Biochem J*. 1991;280(pt 1):267–271.
109. Ratnam M, Marquardt H, Duhning JL, et al. Homologous membrane folate binding proteins in human placenta: cloning and sequence of a cDNA. *Biochemistry*. 1989;28:8249–8254.
110. da Costa M, Rothenberg SP. Purification and characterization of folate binding proteins from rat placenta. *Biochim Biophys Acta*. 1996;1292:23–30.
111. Birn H, Nexø E, Christensen EI, et al. Diversity in rat tissue accumulation of vitamin B₁₂ supports a distinct role for the kidney in vitamin B₁₂ homeostasis. *Nephrol Dial Transplant*. 2003;18:1095–1100.
112. Björn-Yoshimoto WE, Underhill SM. The importance of the excitatory amino acid transporter 3 (EAAT3). *Neurochem Int*. 2016;98:4–18.
113. Drake AJ, O'Shaughnessy PJ, Bhattacharya S, et al. *In utero* exposure to cigarette chemicals induces sex-specific disruption of one-carbon metabolism and DNA methylation in the human fetal liver. *BMC Med*. 2015;13:18. doi:10.1186/s12916-014-0251-x
114. Chen C-S, Kuo Y-T, Tsai H-Y, et al. Brain biochemical correlates of the plasma homocysteine level: a proton magnetic resonance spectroscopy study in the elderly subjects. *Am J Geriatr Psychiatry*. 2011;19:618–626.
115. Barua S, Kuizon S, Ted Brown W, et al. High gestational folic acid supplementation alters expression of imprinted and candidate autism susceptibility genes in a sex-specific manner in mouse offspring. *J Mol Neurosci*. 2016;58:277–286.
116. Kezurer N, Galron D, Golan HM. Increased susceptibility to mild neonatal stress in MTHFR deficient mice. *Behav Brain Res*. 2013;253:240–252.
117. Blumkin E, Levav-Rabkin T, Melamed O, et al. Gender-specific effect of *Mthfr* genotype and neonatal vigabatrin interaction on synaptic proteins in mouse cortex. *Neuropsychopharmacology*. 2011;36:1714–1728.
118. Nugent BM, Wright CL, Shetty AC, et al. Brain feminization requires active repression of masculinization via DNA methylation. *Nat Neurosci*. 2015;18:690–697.
119. Wright EC, Johnson SA, Hao R, et al. Exposure to extrinsic stressors, social defeat or bisphenol A, eliminates sex differences in DNA methyltransferase expression in the amygdala. *J Neuroendocrinol*. 2017;29:1–9.
120. Kigar SL, Chang L, Hayne MR, et al. Sex differences in Gadd45b expression and methylation in the developing rodent amygdala. *Brain Res*. 2016;1642:461–466.
121. Blaze J, Scheuing L, Roth TL. Differential methylation of genes in the medial prefrontal cortex of developing and adult rats following exposure to maltreatment or nurturing care during infancy. *Dev Neurosci*. 2013;35:306–316.
122. Zagni E, Simoni L, Colombo D. Sex and gender differences in central nervous system-related disorders. *Neurosci J*. 2016;2016:1–13.
123. Lenz KM, Nugent BM, McCarthy MM. Sexual differentiation of the rodent brain: dogma and beyond. *Front Neurosci*. 2012;6:1–13.
124. Raznahan A, Shaw P, Lalonde F, et al. How does your cortex grow? *J Neurosci*. 2011;31:7174–7177.
125. Raznahan A, Toro R, Daly E, et al. Cortical anatomy in autism spectrum disorder: an in vivo MRI study on the effect of age. *Cereb Cortex*. 2010;20:1332–1340.

126. McCarthy MM, Auger AP, Bale TL, et al. The epigenetics of sex differences in the brain. *J Neurosci*. 2009;29:12815–12823.
127. Frye CA. Estrus-associated decrements in a water maze task are limited to acquisition. *Physiol Behav*. 1995;57:5–14.
128. Korol DL, Malin EL, Borden KA, et al. Shifts in preferred learning strategy across the estrous cycle in female rats. *Horm Behav*. 2004;45:330–338.
129. Stackman RW, Blasberg ME, Langan CJ, et al. Stability of spatial working memory across the estrous cycle of Long-Evans rats. *Neurobiol Learn Mem*. 1997;67:167–171.
130. Conrad CD, Jackson JL, Wieczorek L, et al. Acute stress impairs spatial memory in male but not female rats: influence of estrous cycle. *Pharmacol Biochem Behav*. 2004;78:569–579.
131. Kolb B, Mychasiuk R, Muhammad A, et al. Experience and the developing prefrontal cortex. *Proc Natl Acad Sci USA*. 2012;109:17186–17193.
132. Auger AP, Tetel MJ, McCarthy MM. Steroid receptor coactivator-1 (SRC-1) mediates the development of sex-specific brain morphology and behavior. *Proc Natl Acad Sci USA*. 2000;97:7551–7555.
133. McCarthy MM, Arnold AP. Reframing sexual differentiation of the brain. *Nat Neurosci*. 2011;14:677–683.